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[54] **DETECTION OF BOTULINUM TOXIN**
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[58] **Field of Search** 536/23.1, 23.53; 435/69.1, 69.6, 70.1, 70.21, 340, 326, 328, 320.1; 935/22, 66-75; 530/387.1, 388.4, 359.5, 866

[56] **References Cited**
FOREIGN PATENT DOCUMENTS
15982 6/1995 WIPO .
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[57] **ABSTRACT**
Recombinant antibody fragments (rFabs) specific to botulinum types A and B toxin complexes are provided. These rFabs are useful as immunosensors for detecting botulinum toxin in food, health care, and military applications. The DNA sequences coding for the rFabs are also disclosed.

31 Claims, No Drawings

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DETECTION OF BOTULINUM TOXIN

This application is a nonprovisional continuation of provisional application Ser. No. 60/011,013, filed Feb. 1, 1996.

GOVERNMENTAL INTEREST

The invention described herein may be manufactured, used and licensed by or for the U.S. Government.

FIELD OF THE INVENTION

The present invention relates generally to the detection of *Clostridium botulinum*. More specifically, the present invention relates to recombinant antibody fragments (rFabs) specific to botulinum types A and B toxin complexes. These rFabs are useful as immunosensors for detecting botulinum toxin in food, health care, and military applications, and may have therapeutic potential.

BACKGROUND OF THE INVENTION

The sole cause of flaccid paralysis in the food poisoning disease known as botulism is a neurotoxin (NT) of approximately 150 kDa, which is produced as antigenically distinct serotypes (types A, B, C, D, E, F and G) by certain strains of *Clostridium botulinum*, *C. butyricum*, and *C. barrati*. In contaminated food, as well as in bacterial culture, the NT is noncovalently associated with non-neurotoxic proteins (non-NTs) in a large, up to 900 kDa, and stable complex. These non-NTs may and may not have hemagglutinating activity and are often immunologically related. For example, type A non-NTs have potent hemagglutinating activity and the non-NTs from type A and B complexes are serologically cross-reactive.

Botulinum toxin complexes, primarily a mixture of types A and B, are also used in biological weapons. Because of the harmful effect such weapons could have on the health of military personnel and civilian populations, as well as the potential for contracting food poisoning from botulinum-contaminated food, there is a need for quick and inexpensive detection of the presence of botulinum toxin type A and/or B in air, water, and food samples.

SUMMARY OF THE INVENTION

Accordingly, it is an object of the present invention to provide an immunosensor for the detection of biological weapons grade botulinum toxin.

Another object of the present invention is to provide an immunosensor that is specific for the non-NT, or hemagglutinating, components of both type A and B toxoids.

Another object of the present invention is to provide such an immunosensor which is rapidly and economically produced.

These and other objects are satisfied by the present invention which is directed to anti-type A and B non-NT rFab fragments (hereinafter anti-AB rFabs) expressed by recombinant bacteria and the use of these anti-AB rFabs for detecting botulinum toxin type A and/or B in air, water, and food samples. The recombinant bacteria were isolated from a recombinant antibody library made using known techniques.

In brief, a cDNA library was made from mouse messenger RNA (mRNA) isolated from the spleens of mice immunized with human pentavalent toxoid emulsified in FCA. The cloned mouse heavy and light chain genes were expressed in

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Escherichia coli phage display libraries as heavy and light chain polypeptides which associate together to form rFab antibody fragments. This combinatorial library was screened against botulinum type B complex to enrich the library for non-NT specific clones. The clones which produced anti-AB rFabs of the invention were then isolated from the enriched library and the DNA sequences of the light and heavy chain coding regions were determined.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides anti-AB rFabs, amino acid sequences thereof, nucleotide sequences coding for said amino acid sequences, recombinant expression vectors containing said nucleotide sequences, and recombinant microorganisms containing said vectors. The anti-AB rFabs of the invention were isolated as described in detail below.

EXAMPLE 1

Toxin Preparations

Botulinum toxin type A complex (500 kDa) and type B complex (500 kDa) were purchased from WAKO BioProducts (Richmond, Va.). The type A and type B neurotoxins (each 150 kDa) were isolated from liquid bacterial culture according to published methods (Sathyamoorthy, V. and DasGupta, B. R., Separation, purification, partial characterization and comparison of the heavy and light chains of botulinum neurotoxins types A, B, and E, J. of Biol. Chem., Vol. 260 (19), pp. 10461-10466 (1985), incorporated herein by reference). The isolated NTs were judged pure by SDS-PAGE and amino acid sequence determinations. Toxoid of the type B complex was prepared as reported elsewhere. (Sugiyama, H., DasGupta, B. R. and Ohishi, I. Disulfide-immunogenicity relationship of botulinum toxins., Proc. of the Soc. for Exper. Biol. and Med., Vol. 145, pp. 1306-1309 (1974), incorporated herein by reference).

EXAMPLE 2

Mouse Immunization Protocol for Recombinant Antibodies

Five, female BALB/c mice were immunized subcutaneously with 0.25 ml of pentavalent botulinum toxoid types A, B, C, D, and E (Michigan Department of Public Health), emulsified in 1 part by volume Complete Freund's Adjuvant (CFA, Difco, Detroit, Mich.). The mice were subsequently boosted three times at 17 day intervals with 0.25 ml of pentavalent vaccine emulsified in 1 part Incomplete Freund's Adjuvant (IFA). Five days after the last immunization, the mice were immunized with 50 μ g of toxoid of botulinum type B complex in IFA. The mice were then primed with an intravenous injection of 100 μ l of botulinum toxin B complex at 2 ng/ml, which also contained 100 μ g of botulinum toxoid type B complex. Mice were killed and spleens removed 3 days after priming immunization. Sera from these animals were pooled and analyzed by direct ELISA, described below, for titer to botulinum toxin type A and B complexes, with an endpoint titer of 1:100,000.

EXAMPLE 3

Anti-Botulinum Toxin Direct ELISA

Direct ELISA assays were performed using 96 well microtiter plates (Immulon II, Dynatech, Chantilly, Va.). The plates were coated overnight at 4° C. with 100 μ l of specified

botulinum toxin at 2 $\mu\text{g}/\text{ml}$ of phosphate buffered saline (PBS 0.01M phosphate buffer, 0.15M NaCl, pH 7.4, Sigma, St. Louis, Mo.) and with bovine serum albumin at the same concentration as the negative antigen. Botulinum type B toxin complex or botulinum type B NT were used as antigens in assays to characterize the specificity of recombinant Fabs. The plates were then washed 6 times using an automated plate washer with PBS Wash Buffer (PBS, 0.1% Tween 20, 0.1% thimerosal, pH=7.4).

Sera diluted in ELISA Dilution Buffer (5% Dry Skim Milk, 0.1% Tween 20, 0.001% thimerosal, PBS pH=7.4) was added to alternating wells of the ELISA plates containing specified botulinum toxin and negative BSA antigen and were incubated for 1 h at 37° C. The plates were then washed again as before with ELISA wash buffer.

Horse radish peroxidase (HRP)-conjugated goat anti-mouse (Kirkegaard and Perry Laboratories, Inc., Gaithersburg, Md.) or anti-Fab (Accurate Chemical & Scientific Corp., Westbury, N.Y.) antibody diluted 1:2500 in ELISA Dilution buffer was added to each well of the ELISA plates and incubated for one hour at 37° C. The plates were washed again 6 times with PBS wash buffer.

ABTS substrate (Kirkegaard and Perry Laboratories, Inc.) was added to each well and incubated for one hour at 37° C. The optical density (OD) at 405 nm of each well of the ELISA plate was determined using an ELISA plate reader (Dynatech). An adjusted OD was obtained by subtracting the OD of the reaction of the antibody or Fab with the negative antigen (BSA) from the OD of the reaction of the antibody solution to the positive antigen. The positive cutoff for this assay was calculated from the adjusted OD of the mean plus three standard deviations of three negative controls consisting of culture media without Fab or monoclonal antibodies or botulinum antibody negative mouse sera.

The polyclonal serum was shown to be responsive towards botulinum toxin complex serotypes A and B (>500 kDa), with an endpoint titer of 1:100,000, as well as towards pure botulinum NT B (150 kDa), that lacked the hemagglutinin commonly found associated with the toxin complex. From the ELISA data, it was concluded that the challenged mice had raised an immune response directed against the 150 kDa NT B as well as towards the non-NT proteins of the toxin complex.

EXAMPLE 4

First Strand cDNA Synthesis and PCR Amplification

Total RNA was isolated from the spleens of a second set of mice immunized as described in Example 2. The isolation procedure was essentially that described by Chomczynski, P. and Sacchi, N., Single step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction, *Anal. Biochem.*, Vol. 162, pp. 156–159 (1987), incorporated herein by reference. Twenty micrograms of the isolated RNA was allowed to anneal with either oligo-dT₁₈ or heavy chain specific immunoglobulin primers, and then extended with 80 units of RNase H⁻ MMLV reverse transcriptase (Stratagene, Inc., LaJolla, Calif.) and 1 mM dNTPs. The resulting cDNAs were processed and amplified essentially as described by Hogrefe, H. and Shopes, B., Construction of Phagemid Display Libraries with PCR-amplified Immunoglobulin Sequences, *PCR Methods Appl.* S109–S122 (1994) (incorporated herein by reference), in order to isolate individual sets of immunoglobulin genes by PCR amplification.

EXAMPLE 5

Construction of Primary Lambda Library

Heavy and light chain PCR fragments were subjected to digestion with Sfi I overnight at 50° C. The digested chains

were gel purified and 1 μg of each set were ligated together in a final volume of 10 μl containing 5 units T4 DNA ligase (Life Technologies, Inc., Gaithersburg, Md.). Two identical ligations were pooled, phenol extracted, ether extracted, and ethanol precipitated. The pellet was resuspended in water and subjected to digestion with 100 units each of Spe I and Not I (Boehringer-Mannheim, Indianapolis, Ind.) overnight at 37° C. The resulting mixture of 1.4 kb inserts was gel isolated in 10 μl of water. Forty ng of the Fab gene insert mixture was ligated into Not I/Spe I digested SurfZAP™ λ arms (Stratagene) essentially as described by Amberg, J., Hogrefe, H. H., et al., SuZAP™ Vector: Linking Phenotype to Genotype for Phagemid Display Libraries, *Strategies Mol. Biol.*, Vol. 5, pp. 2–5 and Vol. 6 (1), pp. 2–4, each of which is incorporated herein by reference. See also Hogrefe, H., Mullinax, R., Lovejoy, A., Hay, B., and Sorge, J., A bacteriophage lambda vector for the cloning and expression of immunoglobulin Fab fragments on the surface of filamentous phage, *Gene* Vol. 128, pp. 119–126 (1993); Hogrefe, H., Amberg, J., Hay, B., Sorge, J. and Shopes, B., Cloning in bacteriophage lambda vector for the display of binding proteins on filamentous phage, *Gene*, Vol. 137, pp. 85–91 (1993), each of which is incorporated herein by reference. The recombinant arms were then packaged into lambda virus (Kretz, P. L., Effect of lambda packaging extract mcr restriction activity on DNA cloning, *Nucleic Acids Res.*, Vol. 17 (13), p. 5409 (1989) incorporated herein by reference). The primary lambda library was titered on XL-1 Blue cells and then amplified using the plate lysate amplification technique (Sambrook, J., Fritsch, E., et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press (1989), incorporated herein by reference).

EXAMPLE 6

Production of Phage for Bio-Panning

The resulting primary lambda library was then subjected to mass excision by infection with ExAssist™ helper phage (Stratagene) in the presence of XL-1 Blue cells (Hay, B. and Short, J. M., ExAssist™ helper phage and SOLR™ cells for Lambda ZAP® II excisions, *Strategies Molec. Biol.*, Vol. 5 (1), pp. 16–18 (1992), herein incorporated by reference). The excised double stranded recombinant pSurfscript™ phagemid were amplified in SOLR™ cells in LB broth, containing 100 $\mu\text{g}/\text{ml}$ carbenicillin and 50 $\mu\text{g}/\text{ml}$ kanamycin (LB^{Carb/Kan} broth), and allowed to amplify for 6 hours at 37° C.

The cells were harvested, resuspended in 10 mM MgSO₄, and 1 \times 10⁹ SOLR™ cells were mixed with 9 \times 10⁹ pfu of VSCM13 helper phage for 15 min. at 37° C. The culture was then diluted to an OD₆₀₀=0.1 with LB^{Carb/Kan} broth and shaken at 30° C. overnight. The culture was centrifuged and the supernatant containing the bacteriophage was precipitated with polyethylene glycol 8000 (PEG). (See McCafferty, J., Griffiths, A. D., et al., Phage Antibodies: filamentous phage displaying antibody variable domains, *Nature*, Vol. 348, pp. 552–554 (1990), incorporated herein by reference). The pellet was resuspended in 1 ml of TE and reprecipitated with PEG. The resulting bacteriophage pellet was resuspended in 100 μl of TE/0.1% BSA.

Co-infection of the cells with the VSCM13 helper virus induced production of filamentous bacteriophage that express the heavy and light chain polypeptides on the surface of the virus. (See Parmley, S. F. and Smith, G. P., Antibody-selectable filamentous fd phage vectors: affinity purification of target genes, *Gene* 73, 305–318 (1988),

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incorporated herein by reference). The display of the rFabs on the surface of the phage allowed for the enrichment of antigen specific clones through biopanning against botulinum type B complex or type B NT immobilized in the wells of microtiter plates as described below.

EXAMPLE 7

Bio-Panning Enrichment and Rescue of Positive Clones

The biopanning procedure used was a modification of two protocols (Persson, M., Caothien, R., and Burton, D., Generation of diverse high affinity human monoclonal antibodies by repertoire cloning, Proc. Natl. Acad. Sci., Vol. 88, pp. 2432-2436 (1991), incorporated herein by reference, and Hogrefe and Shopes (1994)). The wells of Nunc 96 well Immuno-plates were coated for 2 hours at 4° C. with 100 μ l of 10 μ g/ml botulinum type B complex or type B NT in 100 mM sodium bicarbonate buffer, pH=9.0. The plates were then blocked with 1% BSA, 0.5% Tween-20, in PBS (blocking buffer) for 30 minutes at 25° C., followed by three washes with 0.5% Tween-20/PBS (wash buffer).

Approximately 2×10^{10} cfu phagemid were added to each well and the wells were then incubated for 2 hours at 4° C. Unbound phagemids were removed by washing once with wash buffer. To avoid the loss of rare clones the first round of biopanning was washed only once and subsequent rounds were washed two and ten times. The bound phagemids were eluted by adding 100 μ l of 100 mM glycine/0.5% BSA, pH=2.5, to the well and incubating for 10 minutes at 25° C. The elution mix was then mixed briefly and neutralized with 2M Tris base, pH=12.

Eluted phage were used to infect XL-1 Blue cells and the sample was diluted with LB^{carb} and shaken for 1 h at 37° C. VCSM13 helper phage (1×10^{10} pfu) and 50 μ g/ml kanamycin were added and the culture shaken overnight at 29° C. The phagemids were collected as above and the biopanning was repeated for a total of three rounds of selection.

EXAMPLE 8

Colony Lift Hybridization

Following the final round of enrichment, the positive colonies were duplicate plated on LB^{carb} plates, using wetted nitrocellulose filters overlaid onto the bacterial plates. One set of colonies growing directly on the nitrocellulose filters was lysed in a sealed chamber containing a ¼ inch layer of chloroform on the bottom. The filters were submerged in lysozyme buffer (50 mM Tris, pH=8.0, 150 mM NaCl, 5 mM MgCl₂, 3% BSA, 400 μ g/ml lysozyme, and 1 U/ml DNase I). After 1 hour at 25° C., the filters were transferred to a fresh bath of lysozyme for an additional hour, then washed twice for 10 minutes each, in TBST (20 mM Tris, pH=7.5, 150 mM NaCl, 0.05% Tween-20) and blocked for one hour in blocking solution (1% BSA, 20 mM Tris, 150 mM NaCl). Finally, the filters were transferred to 125 ml of blocking solution to which 160 μ l of 200 nM ¹²⁵I-labeled botulinum type B toxin complex (ICN Industries, Irvine, Calif.) or ¹²⁵I-labeled type B NT had been added. (Toxins were labeled using the Chloramine T method and unbound ¹²⁵I was removed by gel filtration on a Sephadex G-10 column run in 0.05M acetic acid, 0.2M NaCl, 0.3% BSA.)

After 1.5 hours at 25° C., the unbound labeled antigen was removed by six 10 minute washes with TBST. The filters were then air dried on blotting paper and exposed to auto-

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radiograph film for 4 hours. Re-alignment of labeled colonies with the master plate allowed the identification of numerous bacterial clones which expressed antibodies capable of binding antigen, i.e., botulinum type B toxin complex or type B NT. Positive clones were subcloned into the pHist purification vector (Tera Biotechnology, LaJolla, Calif.) as described below.

EXAMPLE 9

Subcloning Into pHist and Purification of Fabs

The recombinant pSurfscripTM phagemids were cleaved with Not I/Spe I and the excised Fab gene inserts were recovered in preparative scale in a 2% TAE gel (Amberg, J. et al. (1993); Hogrefe et al. 1994). The purified inserts were then ligated into the pHist vector cut with Not I/Spe I. pHist, a modified version of pSurfscrip, has a linker encoding six histidines inserted in place of the cpIII gene. Bacteria cells (e.g., XL-1 Blue or JM105) transformed with pHist 5 express a rFab comprising a light chain polypeptide and a heavy chain polypeptide fused with a histidine hexapeptide tail. The majority of the expressed rFab is trapped in the periplasmic space with small amounts of the rFab being released into the media upon cell death and lysis.

A Ni-NTA resin (Nickle-Nitrolotriacetate) which binds histidine oligomers with high affinity was used to purify the rFab proteins from sonicated cell extracts of induced clones by affinity chromatography according to the manufacturer (Qiagen, Chatsworth, Calif.). See also Lindner, P., B. Guth, et al., Purification of native proteins from the cytoplasm and periplasm of *Escherichia coli* using IMAC and histidine tails: A comparison of proteins and protocols, Methods: Companion Methods Enzymol. Vol. 4, p. 2, incorporated herein by reference. Antibody protein was eluted from the resin under non-reducing conditions with 500 mM imidazole. This purification with metal chelate affinity chromatography resulted in highly purified rFabs at levels between 500 and 1600 μ g/liter of LB broth (data not shown).

A number of the purified rFabs were analyzed by SDS-PAGE. One clone, pHist5, produced a rFab, named BotFab 5, which migrated as a 50 kDa protein under non-reducing conditions and as two polypeptides of 31 and 29 kDa under reducing conditions (data not shown). Probing a western blot of the SDS-PAGE gel run under non-reducing conditions with mouse anti-Fab antiserum confirmed that the 50 kDa BotFab 5 polypeptide was mouse Fab (data not shown).

EXAMPLE 10

Characterization of Antibody Specificity Towards Botulinum Toxin

Overnight cultures of positive clones were grown and induced with 1 mM isopropyl β -D-thiogalactopyranoside (IPTG) when in early log phase. The cells were pelleted and 100 μ l of the supernatant was allowed to bind for 1 hour to microtiter plates that had been coated with 100 μ l of 5 μ g/ml botulinum type A or B complex, type A or B NT, or bovine serum albumin (BSA) as a negative control. The plates were blocked and washed as described above. The wells were then probed with 100 μ l of goat anti-mouse Fab-HRP conjugate, which served as a secondary calorimetric probe. The conjugate, allowed to bind for 2 hours, was developed after 6 wash cycles by adding ABTS substrate (2'-azino-bis-3-thylbenthiazoline-6-sulphonate) to each well and then incubating for one hour at 37° C. The optical density (OD) at 405 nm of each well of the ELISA plate was determined

using an ELISA plate reader (Dynatech). The results of several representative rFab clones are presented in Table 1 below:

TABLE 1

rFab Name	Type A Complex	Type B Complex	Type A NT	Type B NT	BSA
BotFab 1	1.85	2.70	0.03	0.03	0.33
BotFab 5	0.62	1.40	0.04	0.07	0.05
BotFab 7	2.08	2.80	0.05	0.05	0.05
BotFab 8	1.73	2.40	0.09	0.07	0.09
BotFab 11	0.07	0.07	0.04	0.04	0.04
BotFab 12	1.35	2.05	0.04	0.05	0.06
ToxFab 146	0.03	0.03	0.02	1.41	0.03
ToxFab 149	0.04	0.03	0.02	1.06	0.02
ToxFab 150	0.03	0.03	0.02	0.07	0.03

Data are O.D. units at 405 nm which are not normalized for protein concentration.

As shown by the data in Table 1, rFabs produced by clones isolated after biopanning the λ library against botulinum type B complex, including BotFab 5, were shown by ELISA to be reactive to botulinum types A and B toxin complexes and not reactive to the 150 kDa NT of type A or B. As noted above, the non-NT proteins in the botulinum toxin complexes A and B are immunologically related. Similarly, rFabs produced by clones isolated after biopanning and enriching against the Type B NT were specific to the Type B NT and did not cross-react with Type A NT or the toxin complexes.

Several clones which expressed anti-AB rFabs were selected for DNA sequence analysis. The DNA sequence of the pHist 5 construct is shown in SEQ ID NO:1. The Fab gene insert is separately set forth in SEQ ID NO:2. Nucleotides 117–827 code for the light chain polypeptide and nucleotides 847–1611 code for the heavy chain polypeptide of BotFab 5. Similarly, the DNA sequences of the Fab gene inserts in pHist 1, pHist 20, and pHist 22, which code for BotFab 1, 20, and 22, respectively, are shown in SEQ ID NOS:5, 8, and 11, respectively. The pHist 5 recombinant construct has been deposited with the American Type Culture Collection (Rockville, Md.) as Accession Number 98316 on Feb. 7, 1997. pHist 5 provides the framework for creating other clones. (pHist 1, 20, & 22).

The amino acid sequences of the light and heavy chain polypeptides for each of these constructs were deduced using the DNA STRIDER program (free software available on the Internet) and are shown in SEQ ID NOS:3–4 (BotFab 5), SEQ ID NOS:6–7 (BotFab 1), SEQ ID NOS:9–10 (BotFab 20), and SEQ ID NOS:12–13 (BotFab 22). A comparison of these sequences shows that they contain large stretches of highly homologous regions, suggesting these rFabs all bind to the same non-NT epitope in the types A and B complexes.

Thus, the invention not only includes rFabs containing the recited amino acid sequences of SEQ ID NOS:3–4, 6–7, 9–10, and 12–13, it also includes other rFabs capable of specifically binding to the same epitope. Those rFabs embraced by the invention can be readily determined using commercially available protein folding programs which predict the structure of BotFab 5 and its epitope binding site as well as what changes can be made to its amino acid sequence without changing the binding site. Such rFabs are defined as those having an amino acid sequence with at least 90% homology to the light and heavy chain amino acid sequences of BotFab 5, the preferred rFab of the invention.

The invention also embraces all isolated and purified DNA fragments with DNA sequences coding for the light

and heavy chain amino acid sequences of BotFabs 5, 1, 20, and 22. Thus, the invention includes DNA fragments comprising the coding portions of SEQ ID NOS:2, 5, 8, and 11, or their complementary sequences, and DNA fragments containing DNA sequences which are substantially homologous to the coding portions of SEQ ID NOS:2, 5, 8 and 11 or their complementary sequences.

Substantially homologous DNA sequences are defined as having at least about 85% homology over the defined length of the DNA sequences, with at least about 90% homology being preferred and at least about 95% homology being most preferred. Sequences that are substantially homologous may be identified in a Southern hybridization experiment under stringent conditions. Defining stringent conditions for a particular hybridization experiment is within the skill of the art. Generally, hybridization under stringent conditions is performed at about 5° C. lower than the thermal melting point for the specific sequence at a defined ionic strength and pH (T. Maniatis et al., *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Laboratory (1982), pp. 387–389, incorporated herein by reference). Typical stringent conditions include hybridization in 4×SSC at 65° C. or in 4×SSC, 50% formamide at 42° C., followed by washing in 0.1×SSC at 65° for 30 min.

The DNA fragments according to the invention may be cloned into bacterial expression vectors well-known in the art to generate recombinant expression vectors capable of producing anti-AB rFabs. Preferably, the rFab vectors contain a DNA sequence coding for a purification tail to allow for purification of the expressed rFab by affinity chromatography.

The expression vectors include known transcriptional and translational control elements operatively linked to the light and heavy chain genes. Preferably, such control elements will allow expression of the rFabs in more than one type of microorganism.

The rFabs of the invention, particularly BotFab 5, offer several advantages over monoclonal antibodies for routine testing of biological samples for the presence of botulinum complexes. First, the Fabs are less expensive to produce since they may be isolated from large scale bacterial cultures rather than hybridoma cultures. Also, the affinity of a Fab is more easily altered using known techniques, e.g., by in vitro mutagenesis of its gene and subsequent screening of the expressed Fabs.

The principles, preferred embodiments and modes of operation of the present invention have been described in the foregoing specification. The invention which is intended to be protected herein, however, is not to be construed as limited to the particular forms disclosed, since these are to be regarded as illustrative rather than restrictive. Variations and changes may be made by those skilled in the art without departing from the spirit of the invention. For example, given the sequences disclosed herein, one skilled in the art is capable of combining the light chain coding gene from one pHist construct with the heavy chain coding gene from another phist construct to produce a recombinant construct capable of expressing an anti-AB rFab. Also, nucleotide sequences coding for any known purification tags, including the histidine tail, may be fused to either the light or heavy chain genes.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(iii) NUMBER OF SEQUENCES: 13

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4435 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: circular

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Mus musculus
(B) STRAIN: BALB/c

(vii) IMMEDIATE SOURCE:
(B) CLONE: Clone pHist 5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TTATATACTG ATGCACTTTT CGGGGAAATG TGC GCGGAAC CCCTATTTGT TTATTTTCT	60
AAATACATTC AAATATGTAT CCGCTCATGA GACAATAACC CTGATAAATG CTTCAATAAT	120
ATTGAAAAAG GAAGAGTATG AGTATTCAAC ATTTCCGTGT CGCCCTTATT CCCTTTTTTG	180
CGGCATTTTG CCTTCCTGTT TTGCTCACC CAGAAACGCT GGTGAAAGTA AAAGATGCTG	240
AAGATCAGTT GGGTGCACGA GTGGGTTACA TCGAACTGGA TCTCAACAGC GGTAAGATCC	300
TTGAGAGTTT TCGCCCCGAA GAACGTTTTT CAATGATGAG CACTTTTAAA GTTCTGCTAT	360
GTGGCGCGGT ATTATCCCGT ATTGACGCCG GGCAAGAGCA ACTCGGTTCG CGCATACACT	420
ATTCTCAGAA TGACTTGTTT GAGTACTCAC CAGTCACAGA AAAGCATCTT ACGGATGGCA	480
TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG TGATAACACT GCGGCCAACT	540
TACTTCTGAC AACGATCGGA GGACCGAAGG AGCTAACC GC TTTTTCAC AACATGGGGG	600
ATCATGTAAC TCGCCTTGAT CGTTGGGAAC CGGAGCTGAA TGAAGCCATA CCAAACGACG	660
AGCGTGACAC CACGATGCCT GTAGCAATGG CAACAACGTT GCGCAAATA TTAAGTGGCG	720
AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG GATGGAGGCG GATAAAGTTG	780
CAGGACCACT TCTGCGCTCG GCCCTTCCGG CTGGCTGGTT TATTGCTGAT AAATCTGGAG	840
CCGGTGAGCG TGGGTCTCGG GGTATCATTG CAGCACTGGG GCCAGATGGT AAGCCCTCCC	900
GTATCGTAGT TATCTACACG ACGGGGAGTC AGGCAACTAT GGATGAACGA AATAGACAGA	960
TCGCTGAGAT AGGTGCCTCA CTGATTAAGC ATTGGTAACT GTCAGACCAA GTTTACTCAT	1020
ATATACTTTA GATTGATTTA AACTTCATT TTTAATTTAA AAGGATCTAG GTGAAGATCC	1080
TTTTTGATAA TCTCATGACC AAAATCCCTT AACGTGAGTT TTCGTTCCAC TGAGCGTCAG	1140
ACCCCGTAGA AAAGATCAAA GGATCTTCTT GAGATCCTTT TTTTCTGCGC GTAATCTGCT	1200
GCTTGCAAAC AAAAAACCA CCGCTACCAG CGGTGGTTTG TTTGCCGAT CAAGAGCTAC	1260
CAACTCTTTT TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAAT ACTGTCCTTC	1320
TAGTGTAGCG GTAGTTAGGC CACCACTTCA AGAACTCTGT AGCACC GCCT ACATACCTCG	1380
CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGGCGA TAAGTCGTGT CTTACCGGCT	1440
TGGACTCAAG ACGATAGTTA CCGGATAAGG CGCAGCGGTC GGGCTGAACG GGGGGTTCGT	1500

-continued

GCACACAGCC	CAGCTTGGAG	CGAACGACCT	ACACCGAACT	GAGATACCTA	CAGCGTGAGC	1560
TATGAGAAAAG	CGCCACGCTT	CCCGAAGGGA	GAAAGGCGGA	CAGGTATCCG	GTAAGCGGCA	1620
GGGTCGGAAC	AGGAGAGCGC	ACGAGGGAGC	TTCCAGGGGG	AAACGCCTGG	TATCTTTATA	1680
GTCTGTGCGG	GTTTCGCCAC	CTCTGACTTG	AGCGTCGATT	TTTGTGATGC	TCGTCAGGGG	1740
GGCGGAGCCT	ATGGAAAAAC	GCCAGCAACG	CGGCCTTTTT	ACGGTTCCTG	GCCTTTTGCT	1800
GGCCTTTTGC	TCACATGTTT	TTTCCTGCGT	TATCCCCTGA	TTCTGTGGAT	AACCGTATTA	1860
CCGCCTTTGA	GTGAGCTGAT	ACCGCTCGCC	GCAGCCGAAC	GACCGAGCGC	AGCGAGTCAG	1920
TGAGCGAGGA	AGCGGAAGAG	CGCCCAATAC	GCAAACCGCC	TCTCCCCGCG	CGTTGGCCGA	1980
TTCATTAATG	CAGCTGGCAC	GACAGGTTTC	CCGACTGGAA	AGCGGGCAGT	GAGCGCAACG	2040
CAATTAATGT	GAGTTAGCTC	ACTCATTAGG	CACCCAGGC	TTTACACTTT	ATGCTTCCGG	2100
CTCGTATGTT	GTGTGAATT	GTGAGCGGAT	AACAATTCA	CACAGGAAAC	AGCTATGACC	2160
ATGATTACGC	CAAGCTCGAA	ATTAACCCCTC	ACTAAAGGGA	ACAAAAGCTG	GAGCTTGAAT	2220
TCTTAACACT	TCGCCAAGGA	GACAGTCATA	ATGAAATACC	TATTGCCTAC	GGCGGCCGCT	2280
GGATTGTTAT	TACTCGTGTC	CCAACCAGCC	ATGGCCGACA	TCCAGATGAC	CCAGTCTCCA	2340
GCCTCCCTAT	CTGCATCTGT	GGGAGAAACT	GTCACTATCA	CATGTCGAGC	AAGTGGGAAT	2400
ATTCACAATT	ATTTAGCATG	GTATCAGCAG	AAACAGGGA	AATCTCCTCA	GCTCCTGGTC	2460
TATAATGCAA	AAACCTTAGC	AGATGGTGTG	CCATCAAGGT	TCAGTGGCAG	TGGATCAGGA	2520
ACACAATATT	CTCTCAAGAT	CAACAGCCTG	CAGCCTGAAG	ATTTTGGGAG	TTATTACTGT	2580
CAACATTTTT	GGAGTACTCC	GTGGACGTTT	GGTGGAGGCA	CCAAGCTGGA	AATCAAACGG	2640
GCTGATGCTG	CACCAACTGT	ATCCATCTTC	CCACCATCCA	GTGAGCAGTT	AACATCTGGA	2700
GGTGCCTCAG	TCGTGTGCTT	CTTGAACAAC	TTCTACCCCA	AAGACATCAA	TGTCAAGTGG	2760
AAGATTGATG	GAGTGAAACG	ACAAAATGGC	GTCCTGAACA	GTTGGACTGA	TCAGGACAGC	2820
AAAGACAGCA	CCTACAGCAT	GAGCAGCACC	CTCACATTGA	CCAAGGACGA	GTATGAACGA	2880
CATAACAGCT	ATACCTGTGA	GGCCACTCAC	AAGACATCAA	CTTCACCCAT	TGTCAAGAGC	2940
TTCAACAGGA	ATGAGTGTTA	ATAGCAAGGA	GACAGTCATA	ATGAAATACC	TATTGCCTAC	3000
GGCAGCCGCT	GGATTGTTAT	TACTCGCGGC	CCAACCGGCC	ATGGCCGAGG	TTCAGCTTCA	3060
GCAGTCTGGG	GCAGAGCTTG	TGAAGCCAGG	GGCCTCAGTC	AAGTTGTCCT	GCACAGCTTC	3120
TGGCTTCAAC	ATTAAAGACA	CCTTTATGCA	CTGGGTGAAG	CAGAGGCCTG	AACAGGGCCT	3180
GGAGTGGATT	GGAAGGATTG	ATCCTGCGAA	TGGGAATACT	GAATATGACC	CGAAGTTCCA	3240
GGGCAAGGCC	ACTATAACAG	CAGACACATC	CTCCAACACA	GTCAACCTGC	AGCTCAGCAG	3300
CCTGACATCT	GAGGACACTG	CCGTCTATTA	CTGTGCTAGT	GGAGGGGAAC	TGGGGTTTCC	3360
TTACTGGGGG	CAAGGGACTC	TGGTCACTGT	CTCTGCAGCC	AAAACGACAC	CCCCATCTGT	3420
CTATCCACTG	GCCCCTGGAT	CTGCTGCCCA	AACTAACTCC	ATGGTGACCC	TGGGATGCCT	3480
GGTCAAGGGC	TATTTCCCTG	AGCCAGTGAC	AGTGACCTGG	AACTCTGGAT	CCCTGTCCAG	3540
CGGTGTGCAC	ACCTTCCCAG	CTGTCTTGCA	GTTTGACCTC	TACACTCTGA	GCAGCTCAGT	3600
GACTGTCCCC	TCCAGCACCT	GGCCCAGCGA	GACCGTCACC	TGCAACGTTG	CCCACCCGCG	3660
CAGCAGCACC	AAGTGTGACA	AGAAAATTGT	GCCCAGGGAT	TGTACTAGTG	GAGGTGGAGG	3720
TAGCCACCAT	CACCATCACC	ATTAATCTAG	AGTTAAGCGG	CCGTCGAGGG	GGGGCCCGGT	3780
ACCCAATTCT	CCCTATAGTG	AGTCGTATTA	CAATTCACTG	GCCGTCGTTT	TACAACGTCG	3840
TGACTGGGAA	AACCCTGGCG	TTACCCAACCT	TAATCGCCTT	GCAGCACATC	CCCCTTTCGC	3900

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CAGCTGGCGT AATAGCGAAG AGGCCCGCAC CGATCGCCCT TCCCAACAGT TGC GCAGCCT 3960
GAATGGCGAA TGGAAATGT AAGCGTTAAT ATTTTGTTAA AATTCGCGTT AAATTTTGT 4020
TAAATCAGCT CATTTTTTAA CCAATAGGCC GAAATCGGCA AAATCCCTTA TAAATCAAAA 4080
GAATAGACCG AGATAGGGTT GAGTGTGTGT CCAGTTTGA ACAAGAGTCC ACTATTAAAG 4140
AACGTGGACT CCAACGTCAA AGGGCGAAAA ACCGTCTATC AGGGCGATGG CCCACTACGT 4200
GAACCATCAC CCTAATCAAG TTTTTTGGGG TCGAGGTGCC GTAAAGCACT AAATCGGAAC 4260
CCTAAAGGGA GCCCCGATT TAGAGCTTGA CGGGGAAAGC CGGCGAACGT GCGGAGAAAG 4320
GAAGGGAAGA AAGCGAAAG AGCGGCGCT AGGGCGCTGG CAAGTGTAGC GGTACGCTG 4380
CGCGTAACCA CCACACCCGC CGCGCTTAAT GCGCCGCTAC AGGGCGCGTC AGGTG 4435

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1672 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: Not Relevant
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Mus musculus
 - (B) STRAIN: Balb/c
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: pHist 5
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 117..827
 - (D) OTHER INFORMATION: /product= "antibody fragment, light chain"
/label= BotFab 5
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 847..1611
 - (D) OTHER INFORMATION: /product= "antibody fragment, heavy chain"
/label= BotFab 5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TAGTTATCTC CTTATGTTTC TATTCTCTTT CTCTTACCCA AGCTCGAATT AACCTCACTA 60
AAGGGAACAA AAGCTGGAGC TTGAATCTTT AACTACTCGC CAAGGAGACA GTCATA 116
ATG AAA TAC CTA TTG CCT ACG GCG GCC GCT GGA TTG TTA TTA CTC GCT 164
Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala
1 5 10 15
GCC CAA CCA GCC ATG GCC GAC ATC CAG ATG ACC CAG TCT CCA GCC TCC 212
Ala Gln Pro Ala Met Ala Asp Ile Gln Met Thr Gln Ser Pro Ala Ser
20 25 30
CTA TCT GCA TCT GTG GGA GAA ACT GTC ACT ATC ACA TGT CGA GCA AGT 260
Leu Ser Ala Ser Val Gly Glu Thr Val Thr Ile Thr Cys Arg Ala Ser
35 40 45
GGG AAT ATT CAC AAT TAT TTA GCA TGG TAT CAG CAG AAA CAG GGA AAA 308
Gly Asn Ile His Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Gln Gly Lys
50 55 60
TCT CCT CAG CTC CTG GTC TAT AAT GCA AAA ACC TTA GCA GAT GGT GTG 356
Ser Pro Gln Leu Leu Val Tyr Asn Ala Lys Thr Leu Ala Asp Gly Val
65 70 75 80

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CCA	TCA	AGG	TTC	AGT	GGC	AGT	GGA	TCA	GGA	ACA	CAA	TAT	TCT	CTC	AAG	404
Pro	Ser	Arg	Phe	Ser	Gly	Ser	Gly	Ser	Gly	Thr	Gln	Tyr	Ser	Leu	Lys	
				85					90					95		
ATC	AAC	AGC	CTG	CAG	CCT	GAA	GAT	TTT	GGG	AGT	TAT	TAC	TGT	CAA	CAT	452
Ile	Asn	Ser	Leu	Gln	Pro	Glu	Asp	Phe	Gly	Ser	Tyr	Tyr	Cys	Gln	His	
			100					105					110			
TTT	TGG	AGT	ACT	CCG	TGG	ACG	TTC	GGT	GGA	GGC	ACC	AAG	CTG	GAA	ATC	500
Phe	Trp	Ser	Thr	Pro	Trp	Thr	Phe	Gly	Gly	Gly	Thr	Lys	Leu	Glu	Ile	
		115					120					125				
AAA	CGG	GCT	GAT	GCT	GCA	CCA	ACT	GTA	TCC	ATC	TTC	CCA	CCA	TCC	AGT	548
Lys	Arg	Ala	Asp	Ala	Ala	Pro	Thr	Val	Ser	Ile	Phe	Pro	Pro	Ser	Ser	
	130						135				140					
GAG	CAG	TTA	ACA	TCT	GGA	GGT	GCC	TCA	GTC	GTG	TGC	TTC	TTG	AAC	AAC	596
Glu	Gln	Leu	Thr	Ser	Gly	Gly	Ala	Ser	Val	Val	Cys	Phe	Leu	Asn	Asn	
					150					155				160		
TTC	TAC	CCC	AAA	GAC	ATC	AAT	GTC	AAG	TGG	AAG	ATT	GAT	GGC	AGT	GAA	644
Phe	Tyr	Pro	Lys	Asp	Ile	Asn	Val	Lys	Trp	Lys	Ile	Asp	Gly	Ser	Glu	
				165				170						175		
CGA	CAA	AAT	GGC	GTC	CTG	AAC	AGT	TGG	ACT	GAT	CAG	GAC	AGC	AAA	GAC	692
Arg	Gln	Asn	Gly	Val	Leu	Asn	Ser	Trp	Thr	Asp	Gln	Asp	Ser	Lys	Asp	
			180					185					190			
AGC	ACC	TAC	AGC	ATG	AGC	AGC	ACC	CTC	ACA	TTG	ACC	AAG	GAC	GAG	TAT	740
Ser	Thr	Tyr	Ser	Met	Ser	Ser	Thr	Leu	Thr	Leu	Thr	Lys	Asp	Glu	Tyr	
		195					200					205				
GAA	CGA	CAT	AAC	AGC	TAT	ACC	TGT	GAG	GCC	ACT	CAC	AAG	ACA	TCA	ACT	788
Glu	Arg	His	Asn	Ser	Tyr	Thr	Cys	Glu	Ala	Thr	His	Lys	Thr	Ser	Thr	
		210				215					220					
TCA	CCC	ATT	GTC	AAG	AGC	TTC	AAC	AGG	AAT	GAG	TGT	TAA	TAGCAAGGAG			837
Ser	Pro	Ile	Val	Lys	Ser	Phe	Asn	Arg	Asn	Glu	Cys					
	225				230				235							
ACAGTCATA	ATG	AAA	TAC	CTA	TTG	CCT	ACG	GCA	GCC	GCT	GGA	TTG	TTA			885
	Met	Lys	Tyr	Leu	Leu	Pro	Thr	Ala	Ala	Ala	Gly	Leu	Leu			
		1			5					10						
TTA	CTC	GCG	GCC	CAA	CCG	GCC	ATG	GCC	GAG	GTT	CAG	CTT	CAG	CAG	TCT	933
Leu	Leu	Ala	Ala	Gln	Pro	Ala	Met	Ala	Glu	Val	Gln	Leu	Gln	Gln	Ser	
		15				20					25					
GGG	GCA	GAG	CTT	GTG	AAG	CCA	GGG	GCC	TCA	GTC	AAG	TTG	TCC	TGC	ACA	981
Gly	Ala	Glu	Leu	Val	Lys	Pro	Gly	Ala	Ser	Val	Lys	Leu	Ser	Cys	Thr	
	30				35				40					45		
GCT	TCT	GGC	TTC	AAC	ATT	AAA	GAC	ACC	TTT	ATG	CAC	TGG	GTG	AAG	CAG	1029
Ala	Ser	Gly	Phe	Asn	Ile	Lys	Asp	Thr	Phe	Met	His	Trp	Val	Lys	Gln	
			50						55					60		
AGG	CCT	GAA	CAG	GGC	CTG	GAG	TGG	ATT	GGA	AGG	ATT	GAT	CCT	GCG	AAT	1077
Arg	Pro	Glu	Gln	Gly	Leu	Glu	Trp	Ile	Gly	Arg	Ile	Asp	Pro	Ala	Asn	
			65				70					75				
GGG	AAT	ACT	GAA	TAT	GAC	CCG	AAG	TTC	CAG	GGC	AAG	GCC	ACT	ATA	ACA	1125
Gly	Asn	Thr	Glu	Tyr	Asp	Pro	Lys	Phe	Gln	Gly	Lys	Ala	Thr	Ile	Thr	
		80					85					90				
GCA	GAC	ACA	TCC	TCC	AAC	ACA	GTC	AAC	CTG	CAG	CTC	AGC	AGC	CTG	ACA	1173
Ala	Asp	Thr	Ser	Ser	Asn	Thr	Val	Asn	Leu	Gln	Leu	Ser	Ser	Leu	Thr	
		95				100					105					
TCT	GAG	GAC	ACT	GCC	GTC	TAT	TAC	TGT	GCT	AGT	GGA	GGG	GAA	CTG	GGG	1221
Ser	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala	Ser	Gly	Gly	Glu	Leu	Gly	
	110				115				120					125		
TTT	CCT	TAC	TGG	GGC	CAA	GGG	ACT	CTG	GTC	ACT	GTC	TCT	GCA	GCC	AAA	1269
Phe	Pro	Tyr	Trp	Gly	Gln	Gly	Thr	Leu	Val	Thr	Val	Ser	Ala	Ala	Lys	
			130						135					140		
ACG	ACA	CCC	CCA	TCT	GTC	TAT	CCA	CTG	GCC	CCT	GGA	TCT	GCT	GCC	CAA	1317
Thr	Thr	Pro	Pro	Ser	Val	Tyr	Pro	Leu	Ala	Pro	Gly	Ser	Ala	Ala	Gln	
			145					150						155		

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ACT AAC TCC ATG GTG ACC CTG GGA TGC CTG GTC AAG GGC TAT TTC CCT	1365
Thr Asn Ser Met Val Thr Leu Gly Cys Leu Val Lys Gly Tyr Phe Pro	
160 165 170	
GAG CCA GTG ACA GTG ACC TGG AAC TCT GGA TCC CTG TCC AGC GGT GTG	1413
Glu Pro Val Thr Val Thr Trp Asn Ser Gly Ser Leu Ser Ser Gly Val	
175 180 185	
CAC ACC TTC CCA GCT GTC CTG CAG TTT GAC CTC TAC ACT CTG AGC AGC	1461
His Thr Phe Pro Ala Val Leu Gln Phe Asp Leu Tyr Thr Leu Ser Ser	
190 195 200 205	
TCA GTG ACT GTC CCC TCC AGC ACC TGG CCC AGC GAG ACC GTC ACC TGC	1509
Ser Val Thr Val Pro Ser Ser Thr Trp Pro Ser Glu Thr Val Thr Cys	
210 215 220	
AAC GTT GCC CAC CCG GCC AGC AGC ACC AAG GTG GAC AAG AAA ATT GTG	1557
Asn Val Ala His Pro Ala Ser Ser Thr Lys Val Asp Lys Lys Ile Val	
225 230 235	
CCC AGG GAT TGT ACT AGT GGA GGT GGA GGT AGC CAC CAT CAC CAT CAC	1605
Pro Arg Asp Cys Thr Ser Gly Gly Gly Gly Ser His His His His His	
240 245 250	
CAT TAA TCTAGAGTTA AGCGGCCGTC GAGGGGGGGC CCGGTACCCA ATTCGCCCTA	1661
His	
TAGTGAGTCG T	1672

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 236 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala	
1 5 10 15	
Ala Gln Pro Ala Met Ala Asp Ile Gln Met Thr Gln Ser Pro Ala Ser	
20 25 30	
Leu Ser Ala Ser Val Gly Glu Thr Val Thr Ile Thr Cys Arg Ala Ser	
35 40 45	
Gly Asn Ile His Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Gln Gly Lys	
50 55 60	
Ser Pro Gln Leu Leu Val Tyr Asn Ala Lys Thr Leu Ala Asp Gly Val	
65 70 75 80	
Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Gln Tyr Ser Leu Lys	
85 90 95	
Ile Asn Ser Leu Gln Pro Glu Asp Phe Gly Ser Tyr Tyr Cys Gln His	
100 105 110	
Phe Trp Ser Thr Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile	
115 120 125	
Lys Arg Ala Asp Ala Ala Pro Thr Val Ser Ile Phe Pro Pro Ser Ser	
130 135 140	
Glu Gln Leu Thr Ser Gly Gly Ala Ser Val Val Cys Phe Leu Asn Asn	
145 150 155 160	
Phe Tyr Pro Lys Asp Ile Asn Val Lys Trp Lys Ile Asp Gly Ser Glu	
165 170 175	
Arg Gln Asn Gly Val Leu Asn Ser Trp Thr Asp Gln Asp Ser Lys Asp	
180 185 190	
Ser Thr Tyr Ser Met Ser Ser Thr Leu Thr Leu Thr Lys Asp Glu Tyr	
195 200 205	

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Glu Arg His Asn Ser Tyr Thr Cys Glu Ala Thr His Lys Thr Ser Thr
 210 215 220

Ser Pro Ile Val Lys Ser Phe Asn Arg Asn Glu Cys
 225 230 235

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 254 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala
 1 5 10 15
 Ala Gln Pro Ala Met Ala Glu Val Gln Leu Gln Gln Ser Gly Ala Glu
 20 25 30
 Leu Val Lys Pro Gly Ala Ser Val Lys Leu Ser Cys Thr Ala Ser Gly
 35 40 45
 Phe Asn Ile Lys Asp Thr Phe Met His Trp Val Lys Gln Arg Pro Glu
 50 55 60
 Gln Gly Leu Glu Trp Ile Gly Arg Ile Asp Pro Ala Asn Gly Asn Thr
 65 70 75 80
 Glu Tyr Asp Pro Lys Phe Gln Gly Lys Ala Thr Ile Thr Ala Asp Thr
 85 90 95
 Ser Ser Asn Thr Val Asn Leu Gln Leu Ser Ser Leu Thr Ser Glu Asp
 100 105 110
 Thr Ala Val Tyr Tyr Cys Ala Ser Gly Gly Glu Leu Gly Phe Pro Tyr
 115 120 125
 Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ala Ala Lys Thr Thr Pro
 130 135 140
 Pro Ser Val Tyr Pro Leu Ala Pro Gly Ser Ala Ala Gln Thr Asn Ser
 145 150 155 160
 Met Val Thr Leu Gly Cys Leu Val Lys Gly Tyr Phe Pro Glu Pro Val
 165 170 175
 Thr Val Thr Trp Asn Ser Gly Ser Leu Ser Ser Gly Val His Thr Phe
 180 185 190
 Pro Ala Val Leu Gln Phe Asp Leu Tyr Thr Leu Ser Ser Ser Val Thr
 195 200 205
 Val Pro Ser Ser Thr Trp Pro Ser Glu Thr Val Thr Cys Asn Val Ala
 210 215 220
 His Pro Ala Ser Ser Thr Lys Val Asp Lys Lys Ile Val Pro Arg Asp
 225 230 235 240
 Cys Thr Ser Gly Gly Gly Gly Ser His His His His His
 245 250

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1641 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: Not Relevant

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

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(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Mus musculus
 (B) STRAIN: BALB/c

(vii) IMMEDIATE SOURCE:
 (B) CLONE: pHist 1

(ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 87..788
 (D) OTHER INFORMATION: /product= "antibody fragment, light chain"
 /label= BotFab 1

(ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 815..1579
 (D) OTHER INFORMATION: /product= "antibody fragment, Heavy Chain"
 /label= BotFab 1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATACGCCAAG CTCGAAATTA ACCCTCACTA AAGGGAACAA AAGCTGGAGC TTGAATTCTT	60
AACTACTCGC CAAGGAGACA GTCATA ATG AAA TAC CTA TTG CCT ACG GCG GCC	113
Met Lys Tyr Leu Leu Pro Thr Ala Ala	
1 5	
GCT GGA TTG TTA TTA CTC GCT GCC CAA CCA GCC ATG GCC GAC ATC CAG	161
Ala Gly Leu Leu Leu Leu Ala Ala Gln Pro Ala Met Ala Asp Ile Gln	
10 15 20 25	
ATG ACC CAG TCT CCA GCC TCC CTA TCT GCA TCT GTG GGA GAA ACT GTC	209
Met Thr Gln Ser Pro Ala Ser Leu Ser Ala Ser Val Gly Glu Thr Val	
30 35 40	
ACT ATC ACA TGT CGA GCA AGT GGG AAT ATT CAC AAT TAT TTA GCA TGG	257
Thr Ile Thr Cys Arg Ala Ser Gly Asn Ile His Asn Tyr Leu Ala Trp	
45 50 55	
TAT CAG CAG AAA CAG GGA AAA TCT CCT CAG CTC CTG GTC TAT AAT GCA	305
Tyr Gln Gln Lys Gln Gly Lys Ser Pro Gln Leu Leu Val Tyr Asn Ala	
60 65 70	
AAA ACC TTA GCA GAT GGT GTG CCA TCA AGG TTC AGT GGC AGT GGA TCA	353
Lys Thr Leu Ala Asp Gly Val Pro Ser Arg Phe Ser Gly Ser Gly Ser	
75 80 85	
GGA ACA CAA TAT TCT CTC AAG ATC AAC AGC CTG CAG CCT GAA GAT TTT	401
Gly Thr Gln Tyr Ser Leu Lys Ile Asn Ser Leu Gln Pro Glu Asp Phe	
90 95 100 105	
GGG AGT TAT TAC TGT CAA CAT TTT TGG AGT ACT CCG TGG ACG TTC GGT	449
Gly Ser Tyr Tyr Cys Gln His Phe Trp Ser Thr Pro Trp Thr Phe Gly	
110 115 120	
GGA GGC ACC AAG CTG GAA ATC AAA CGG GCT GAT GCT GCA CCA ACT GTA	497
Gly Gly Thr Lys Leu Glu Ile Lys Arg Ala Asp Ala Ala Pro Thr Val	
125 130 135	
TCC ATC TTC CCA CCA TCC AGT GAG CAG TTA ACA TCT GGA GGT GCC TCA	545
Ser Ile Phe Pro Pro Ser Ser Glu Gln Leu Thr Ser Gly Gly Ala Ser	
140 145 150	
GTC GTG TGC TTC TTG AAC AAC TTC TAC CCC AAA GAC ATC AAT GTC AAG	593
Val Val Cys Phe Leu Asn Asn Phe Tyr Pro Lys Asp Ile Asn Val Lys	
155 160 165	
TGG AAG ATT GAT GGC AGT GAA CGA CAA AAT GGC GTC CTG AAC AGT TGG	641
Trp Lys Ile Asp Gly Ser Glu Arg Gln Asn Gly Val Leu Asn Ser Trp	
170 175 180 185	
ACT GAT CAG GAC AGC AAA GAC AGC ACC TAC AGC ATG AGC AGC ACC CTC	689
Thr Asp Gln Asp Ser Lys Asp Ser Thr Tyr Ser Met Ser Ser Thr Leu	
190 195 200	

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ACA TTG ACC AAG GAC GAG TAT GAA CGA CAT AAC AGC TAT ACC TGT GAG	737
Thr Leu Thr Lys Asp Glu Tyr Glu Arg His Asn Ser Tyr Thr Cys Glu	
205 210 215	
GCC ACT CAC AAG ACA TCA ACT TCC CCA TTG TCT AGA GCT TCC CAG GAA	785
Ala Thr His Lys Thr Ser Thr Ser Pro Leu Ser Arg Ala Ser Gln Glu	
220 225 230	
TGA GTGTTTATAG CAAGGAAACA GTCATA ATG AAA TAC CTA TTG CCT ACG GCA	838
Met Lys Tyr Leu Leu Pro Thr Ala	
1 5	
GCC GCT GGA TTG TTA TTA CTC GCG GCC CAA CCG GCG ATG GCC GAG GTT	886
Ala Ala Gly Leu Leu Leu Leu Ala Ala Gln Pro Ala Met Ala Glu Val	
10 15 20	
CAG CTT CAG CAG TCT GGG GCA GAG CTT GTG AAG CCA GGG GCC TCA GTC	934
Gln Leu Gln Gln Ser Gly Ala Glu Leu Val Lys Pro Gly Ala Ser Val	
25 30 35 40	
AAG TTG TCC TGC ACA GCT TCT GGC TTC AAC ATT AAA GAC ACC TTT ATG	982
Lys Leu Ser Cys Thr Ala Ser Gly Phe Asn Ile Lys Asp Thr Phe Met	
45 50 55	
CAC TGG GTG AAG CAG AGG CCT GAA CAG GGC CTG GAG TGG ATT GGA AGG	1030
His Trp Val Lys Gln Arg Pro Glu Gln Gly Leu Glu Trp Ile Gly Arg	
60 65 70	
ATT GAT CCT GCG AAT GGG AAT ACT GAA TAT GAC CCG AAG TTC CAG GGC	1078
Ile Asp Pro Ala Asn Gly Asn Thr Glu Tyr Asp Pro Lys Phe Gln Gly	
75 80 85	
AAG GCC ACT ATA ACA GCA GAC ACA TCC TCC AAC ACA GTC AAC CTG CAG	1126
Lys Ala Thr Ile Thr Ala Asp Thr Ser Ser Asn Thr Val Asn Leu Gln	
90 95 100	
CTC AGC AGC CTG ACA TCT GAG GAC ACT GCC GTC TAT TAC TGT GCT AGT	1174
Leu Ser Ser Leu Thr Ser Glu Asp Thr Ala Val Tyr Tyr Cys Ala Ser	
105 110 115 120	
GGA GGG GAA CTG GGG TTT CCT TAC TGG GGC CAA GGG ACT CTG GTC ACT	1222
Gly Gly Glu Leu Gly Phe Pro Tyr Trp Gly Gln Gly Thr Leu Val Thr	
125 130 135	
GTC TCT GCA GCC AAA ACG ACA CCC CCA TCT GTC TAT CCA CTG GCC CCT	1270
Val Ser Ala Ala Lys Thr Thr Pro Pro Ser Val Tyr Pro Leu Ala Pro	
140 145 150	
GGA TCT GCT GCC CAA ACT AAC TCC ATG GTG ACC CTG GGA TGC CTG GTC	1318
Gly Ser Ala Ala Gln Thr Asn Ser Met Val Thr Leu Gly Cys Leu Val	
155 160 165	
AAG GGC TAT TTC CCT GAG CCA GTG ACA GTG ACC TGG AAC TCT GGA TCC	1366
Lys Gly Tyr Phe Pro Glu Pro Val Thr Val Thr Trp Asn Ser Gly Ser	
170 175 180	
CTG TCC AGC GGT GTG CAC ACC TTC CCA GCT GTC CTG CAG TAT GAC CTC	1414
Leu Ser Ser Gly Val His Thr Phe Pro Ala Val Leu Gln Tyr Asp Leu	
185 190 195 200	
TAC ACT CTG AGC AGC TCA GTG ACT GTC CCC TCC AGC ACC TGG CCC AGC	1462
Tyr Thr Leu Ser Ser Ser Val Thr Val Pro Ser Ser Thr Trp Pro Ser	
205 210 215	
GAG ACC GTC ACC TGC AAC GTT GCC CAC CCG GCC AGC AGC ACC AAG GTG	1510
Glu Thr Val Thr Cys Asn Val Ala His Pro Ala Ser Ser Thr Lys Val	
220 225 230	
GAC AAG AAA ATT GTG CCC AGG GAT TGT ACT AGT GGA GGT GGA GGT AGC	1558
Asp Lys Lys Ile Val Pro Arg Asp Cys Thr Ser Gly Gly Gly Gly Ser	
235 240 245	
CAC CAT CAC CAT CAC CAT TAA TCTAGAGTTA AGCGGCCGTC GAGGGGGGGC	1609
His His His His His	
250	
CCGGTACCCA ATTCGCCCTA TAGTGAGTCG TA	1641

(2) INFORMATION FOR SEQ ID NO:6:

-continued

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 233 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

```

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala
 1           5           10           15
Ala Gln Pro Ala Met Ala Asp Ile Gln Met Thr Gln Ser Pro Ala Ser
          20           25           30
Leu Ser Ala Ser Val Gly Glu Thr Val Thr Ile Thr Cys Arg Ala Ser
          35           40           45
Gly Asn Ile His Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Gln Gly Lys
 50           55           60
Ser Pro Gln Leu Leu Val Tyr Asn Ala Lys Thr Leu Ala Asp Gly Val
 65           70           75           80
Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Gln Tyr Ser Leu Lys
          85           90           95
Ile Asn Ser Leu Gln Pro Glu Asp Phe Gly Ser Tyr Tyr Cys Gln His
          100          105          110
Phe Trp Ser Thr Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile
          115          120          125
Lys Arg Ala Asp Ala Ala Pro Thr Val Ser Ile Phe Pro Pro Ser Ser
          130          135          140
Glu Gln Leu Thr Ser Gly Gly Ala Ser Val Val Cys Phe Leu Asn Asn
          145          150          155          160
Phe Tyr Pro Lys Asp Ile Asn Val Lys Trp Lys Ile Asp Gly Ser Glu
          165          170          175
Arg Gln Asn Gly Val Leu Asn Ser Trp Thr Asp Gln Asp Ser Lys Asp
          180          185          190
Ser Thr Tyr Ser Met Ser Ser Thr Leu Thr Leu Thr Lys Asp Glu Tyr
          195          200          205
Glu Arg His Asn Ser Tyr Thr Cys Glu Ala Thr His Lys Thr Ser Thr
          210          215          220
Ser Pro Leu Ser Arg Ala Ser Gln Glu
          225          230

```

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 254 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

```

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala
 1           5           10           15
Ala Gln Pro Ala Met Ala Glu Val Gln Leu Gln Gln Ser Gly Ala Glu
          20           25           30
Leu Val Lys Pro Gly Ala Ser Val Lys Leu Ser Cys Thr Ala Ser Gly
          35           40           45
Phe Asn Ile Lys Asp Thr Phe Met His Trp Val Lys Gln Arg Pro Glu
          50           55           60

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Gln	Gly	Leu	Glu	Trp	Ile	Gly	Arg	Ile	Asp	Pro	Ala	Asn	Gly	Asn	Thr	
65					70					75						80
Glu	Tyr	Asp	Pro	Lys	Phe	Gln	Gly	Lys	Ala	Thr	Ile	Thr	Ala	Asp	Thr	
				85					90					95		
Ser	Ser	Asn	Thr	Val	Asn	Leu	Gln	Leu	Ser	Ser	Leu	Thr	Ser	Glu	Asp	
			100					105					110			
Thr	Ala	Val	Tyr	Tyr	Cys	Ala	Ser	Gly	Gly	Glu	Leu	Gly	Phe	Pro	Tyr	
		115					120					125				
Trp	Gly	Gln	Gly	Thr	Leu	Val	Thr	Val	Ser	Ala	Ala	Lys	Thr	Thr	Pro	
	130					135					140					
Pro	Ser	Val	Tyr	Pro	Leu	Ala	Pro	Gly	Ser	Ala	Ala	Gln	Thr	Asn	Ser	
145					150				155						160	
Met	Val	Thr	Leu	Gly	Cys	Leu	Val	Lys	Gly	Tyr	Phe	Pro	Glu	Pro	Val	
			165					170					175			
Thr	Val	Thr	Trp	Asn	Ser	Gly	Ser	Leu	Ser	Ser	Gly	Val	His	Thr	Phe	
		180					185						190			
Pro	Ala	Val	Leu	Gln	Tyr	Asp	Leu	Tyr	Thr	Leu	Ser	Ser	Ser	Val	Thr	
		195					200					205				
Val	Pro	Ser	Ser	Thr	Trp	Pro	Ser	Glu	Thr	Val	Thr	Cys	Asn	Val	Ala	
	210					215					220					
His	Pro	Ala	Ser	Ser	Thr	Lys	Val	Asp	Lys	Lys	Ile	Val	Pro	Arg	Asp	
225					230				235						240	
Cys	Thr	Ser	Gly	Gly	Gly	Gly	Ser	His	His	His	His	His	His	His		
			245					250								

- (2) INFORMATION FOR SEQ ID NO:8:
- (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1632 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: Not Relevant
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:

(A) ORGANISM: Mus musculus

(B) STRAIN: BALB/c
- (vii) IMMEDIATE SOURCE:

(B) CLONE: pHist 20
- (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 88..798

(D) OTHER INFORMATION: /product= "antibody fragment, light chain"

/label= BotFab 20
- (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 818..1582

(D) OTHER INFORMATION: /product= "antibody fragment, Heavy Chain"

/label= BotFab 20
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
- TATACGCCAA GCTCGAAATT AACCCCTCACT AAAGGGAACA AAAGCTGGAG CTTGAATTCT

60
- TAAGTACTCG CCAAGGAGAC AGTCATA ATG AAA TAC CTA TTG CCT ACG GCG

111
- Met Lys Tyr Leu Leu Pro Thr Ala

15

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GCC	GCT	GGA	TTG	TTA	TTA	CTC	GCT	GCC	CAA	CCA	GCC	ATG	GCC	GAC	ATC	159
Ala	Ala	Gly	Leu	Leu	Leu	Leu	Ala	Ala	Gln	Pro	Ala	Met	Ala	Asp	Ile	
	10					15					20					
CAG	ATG	ACC	CAG	TCT	CCA	GCC	TCC	CTA	TCT	GCA	TCT	GTG	GGA	GAA	ACT	207
Gln	Met	Thr	Gln	Ser	Pro	Ala	Ser	Leu	Ser	Ala	Ser	Val	Gly	Glu	Thr	
	25				30					35					40	
GTC	ACT	ATC	ACA	TGT	CGA	GCA	AGT	GGG	AAT	ATT	CAC	AAT	TAT	TTA	GCA	255
Val	Thr	Ile	Thr	Cys	Arg	Ala	Ser	Gly	Asn	Ile	His	Asn	Tyr	Leu	Ala	
				45					50					55		
TGG	TAT	CAG	CAG	AAA	CAG	GGG	AAA	TCT	CCT	CAG	CTC	CTG	GTC	TAT	AAT	303
Trp	Tyr	Gln	Gln	Lys	Gln	Gly	Lys	Ser	Pro	Gln	Leu	Leu	Val	Tyr	Asn	
				60				65					70			
GCA	AAA	ACC	TTA	GCA	GAT	GGT	GTG	CCA	TCA	AGG	TTC	AGT	GGC	AGT	GGA	351
Ala	Lys	Thr	Leu	Ala	Asp	Gly	Val	Pro	Ser	Arg	Phe	Ser	Gly	Ser	Gly	
		75					80					85				
TCA	GGA	ACA	CAA	TAT	TCT	CTC	AAG	ATC	AAC	AGC	CTG	CAG	CCT	GAA	GAT	399
Ser	Gly	Thr	Gln	Tyr	Ser	Leu	Lys	Ile	Asn	Ser	Leu	Gln	Pro	Glu	Asp	
	90					95					100					
TTT	GGG	AGT	TAT	TAC	TGT	CAA	CAT	TTT	TGG	AGT	ACT	CCG	TGG	ACG	TTC	447
Phe	Gly	Ser	Tyr	Tyr	Cys	Gln	His	Phe	Trp	Ser	Thr	Pro	Trp	Thr	Phe	
	105				110					115					120	
GGT	GGA	GGC	ACC	AAG	CTG	GAA	ATC	AAA	CGG	GCT	GAT	GCT	GCA	CCA	ACT	495
Gly	Gly	Gly	Thr	Lys	Leu	Glu	Ile	Lys	Arg	Ala	Asp	Ala	Ala	Pro	Thr	
				125					130					135		
GTA	TCC	ATC	TTC	CCA	CCA	TCC	AGT	GAG	CAG	TTA	ACA	TCT	GGA	GGT	GCC	543
Val	Ser	Ile	Phe	Pro	Pro	Ser	Ser	Glu	Gln	Leu	Thr	Ser	Gly	Gly	Ala	
			140					145					150			
TCA	GTC	GTG	TGC	TTC	TTG	AAC	AAC	TTC	TAC	CCC	AAA	GAC	ATC	AAT	GTC	591
Ser	Val	Val	Cys	Phe	Leu	Asn	Asn	Phe	Tyr	Pro	Lys	Asp	Ile	Asn	Val	
		155				160						165				
AAG	TGG	AAG	ATT	GAT	GGC	AGT	GAA	CGA	CAA	AAT	GGC	GTC	CTG	AAC	AGT	639
Lys	Trp	Lys	Ile	Asp	Gly	Ser	Glu	Arg	Gln	Asn	Gly	Val	Leu	Asn	Ser	
	170					175					180					
TGG	ACT	GAT	CAG	GAC	AGC	AAA	GAC	AGC	ACC	TAC	AGC	ATG	AGC	AGC	ACC	687
Trp	Thr	Asp	Gln	Asp	Ser	Lys	Asp	Ser	Thr	Tyr	Ser	Met	Ser	Ser	Thr	
	185				190					195					200	
CTC	ACA	TTG	ACC	AAG	GAC	GAG	TAT	GAA	CGA	CAT	AAC	AGC	TAT	ACC	TGT	735
Leu	Thr	Leu	Thr	Lys	Asp	Glu	Tyr	Glu	Arg	His	Asn	Ser	Tyr	Thr	Cys	
				205					210					215		
GAG	GCC	ACT	CAC	AAG	ACT	TCA	ACT	TCA	CCC	ATT	GTC	AAG	AGC	TTC	AAC	783
Glu	Ala	Thr	His	Lys	Thr	Ser	Thr	Ser	Pro	Ile	Val	Lys	Ser	Phe	Asn	
			220					225					230			
AGG	AAT	GAG	TGT	TAA	TAGCAAGGAG	ACAGTCATA	ATG	AAA	TAC	CTA	TTG	CCT				835
Arg	Asn	Glu	Cys					Met	Lys	Tyr	Leu	Leu	Pro			
	235							1			5					
ACG	GCA	GCC	GTT	GGA	TTG	TTA	TTA	CTC	GCG	GCC	CAA	CCG	GCC	ATG	GCC	883
Thr	Ala	Ala	Val	Gly	Leu	Leu	Leu	Leu	Ala	Ala	Gln	Pro	Ala	Met	Ala	
			10					15					20			
GAG	GTT	CAG	CTT	CAG	CAG	TCT	GGG	GCA	GAG	CTT	GTG	AAG	CCA	GGG	GCC	931
Glu	Val	Gln	Leu	Gln	Gln	Ser	Gly	Ala	Glu	Leu	Val	Lys	Pro	Gly	Ala	
	25					30						35				
TCA	GTC	AAG	TTG	TCC	TGC	ACA	GCT	TCT	GGC	TTC	AAC	ATT	AAA	GAC	ACC	979
Ser	Val	Lys	Leu	Ser	Cys	Thr	Ala	Ser	Gly	Phe	Asn	Ile	Lys	Asp	Thr	
	40					45					50					
TTT	ATG	CAC	TGG	GTG	AAG	CAG	AGG	CCT	GAA	CAG	GGC	CTG	GAG	TGG	ATT	1027
Phe	Met	His	Trp	Val	Lys	Gln	Arg	Pro	Glu	Gln	Gly	Leu	Glu	Trp	Ile	
	55				60				65						70	
GGA	AGG	ATT	GAT	CCT	GCG	AAT	GGG	AAT	ACT	GAA	TAT	GAC	CCG	AAG	TTC	1075
Gly	Arg	Ile	Asp	Pro	Ala	Asn	Gly	Asn	Thr	Glu	Tyr	Asp	Pro	Lys	Phe	
			75					80						85		

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CAG GGC AAG GCC ACT ATA ACA GCA GAC ACA TCC TCC AAC ACA GTC AAC	1123
Gln Gly Lys Ala Thr Ile Thr Ala Asp Thr Ser Ser Asn Thr Val Asn	
90 95 100	
CTG CAG CTC AGC AGC CTG ACA TCT GAG GAC ACT GCC GTC TAT TAC TGT	1171
Leu Gln Leu Ser Ser Leu Thr Ser Glu Asp Thr Ala Val Tyr Tyr Cys	
105 110 115	
GCT AGT GGA GGG GAA CTG GGG TTT CCT TAC TGG GGC CAA GGG ACT CTG	1219
Ala Ser Gly Gly Glu Leu Gly Phe Pro Tyr Trp Gly Gln Gly Thr Leu	
120 125 130	
GTC ACT GTC TCT GCA GCC AAA ACG ACA CCC CCA TCT GTC TAT CCA CTG	1267
Val Thr Val Ser Ala Ala Lys Thr Thr Pro Pro Ser Val Tyr Pro Leu	
135 140 145 150	
GCC CCT GGA TCT GCT GCC CAA ACT AAC TCC ATG GTG ACC CTG GGA TGC	1315
Ala Pro Gly Ser Ala Ala Gln Thr Asn Ser Met Val Thr Leu Gly Cys	
155 160 165	
CTG GTC AAG GGC TAT TTC CCT GAG CCA GTG ACA GTG ACC TGG AAC TCT	1363
Leu Val Lys Gly Tyr Phe Pro Glu Pro Val Thr Val Thr Trp Asn Ser	
170 175 180	
GGA TCC CTG TCC AGC GGT GTG CAC ACC TTC CCA GCT GTC CTG CAG TCT	1411
Gly Ser Leu Ser Ser Gly Val His Thr Phe Pro Ala Val Leu Gln Ser	
185 190 195	
GAC CTC TAC ACT CTG AGC AGC TCA GTG ACT GTC CCC TCC AGC ACC TGG	1459
Asp Leu Tyr Thr Leu Ser Ser Ser Val Thr Val Pro Ser Ser Thr Trp	
200 205 210	
CCC AGC GAG ACC GTC ACC TGC AAC GTT GCC CAC CCG GCC AGC AGC ACC	1507
Pro Ser Glu Thr Val Thr Cys Asn Val Ala His Pro Ala Ser Ser Thr	
215 220 225 230	
AAG GTG GAC AAG AAA ATT GTG CCC AGG GAT TGT ACT AGT GGA GGT GGA	1555
Lys Val Asp Lys Lys Ile Val Pro Arg Asp Cys Thr Ser Gly Gly Gly	
235 240 245	
GGT AGC CAC CAT CAC CAT CAC CAT TAA TCTAGAGTTA AGCGGCCGTC	1602
Gly Ser His His His His His	
250	
GAGGGGGCCC CGATACCCAA TTCGCCTTAT	1632

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 236 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Ala	
1 5 10 15	
Ala Gln Pro Ala Met Ala Asp Ile Gln Met Thr Gln Ser Pro Ala Ser	
20 25 30	
Leu Ser Ala Ser Val Gly Glu Thr Val Thr Ile Thr Cys Arg Ala Ser	
35 40 45	
Gly Asn Ile His Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Gln Gly Lys	
50 55 60	
Ser Pro Gln Leu Leu Val Tyr Asn Ala Lys Thr Leu Ala Asp Gly Val	
65 70 75 80	
Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Gln Tyr Ser Leu Lys	
85 90 95	
Ile Asn Ser Leu Gln Pro Glu Asp Phe Gly Ser Tyr Tyr Cys Gln His	
100 105 110	

-continued

Phe Trp Ser Thr Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile
 115 120 125
 Lys Arg Ala Asp Ala Ala Pro Thr Val Ser Ile Phe Pro Pro Ser Ser
 130 135 140
 Glu Gln Leu Thr Ser Gly Gly Ala Ser Val Val Cys Phe Leu Asn Asn
 145 150 155 160
 Phe Tyr Pro Lys Asp Ile Asn Val Lys Trp Lys Ile Asp Gly Ser Glu
 165 170 175
 Arg Gln Asn Gly Val Leu Asn Ser Trp Thr Asp Gln Asp Ser Lys Asp
 180 185 190
 Ser Thr Tyr Ser Met Ser Ser Thr Leu Thr Leu Thr Lys Asp Glu Tyr
 195 200 205
 Glu Arg His Asn Ser Tyr Thr Cys Glu Ala Thr His Lys Thr Ser Thr
 210 215 220
 Ser Pro Ile Val Lys Ser Phe Asn Arg Asn Glu Cys
 225 230 235

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 254 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Lys Tyr Leu Leu Pro Thr Ala Ala Val Gly Leu Leu Leu Leu Ala
 1 5 10 15
 Ala Gln Pro Ala Met Ala Glu Val Gln Leu Gln Gln Ser Gly Ala Glu
 20 25 30
 Leu Val Lys Pro Gly Ala Ser Val Lys Leu Ser Cys Thr Ala Ser Gly
 35 40 45
 Phe Asn Ile Lys Asp Thr Phe Met His Trp Val Lys Gln Arg Pro Glu
 50 55 60
 Gln Gly Leu Glu Trp Ile Gly Arg Ile Asp Pro Ala Asn Gly Asn Thr
 65 70 75 80
 Glu Tyr Asp Pro Lys Phe Gln Gly Lys Ala Thr Ile Thr Ala Asp Thr
 85 90 95
 Ser Ser Asn Thr Val Asn Leu Gln Leu Ser Ser Leu Thr Ser Glu Asp
 100 105 110
 Thr Ala Val Tyr Tyr Cys Ala Ser Gly Gly Glu Leu Gly Phe Pro Tyr
 115 120 125
 Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ala Ala Lys Thr Thr Pro
 130 135 140
 Pro Ser Val Tyr Pro Leu Ala Pro Gly Ser Ala Ala Gln Thr Asn Ser
 145 150 155 160
 Met Val Thr Leu Gly Cys Leu Val Lys Gly Tyr Phe Pro Glu Pro Val
 165 170 175
 Thr Val Thr Trp Asn Ser Gly Ser Leu Ser Ser Gly Val His Thr Phe
 180 185 190
 Pro Ala Val Leu Gln Ser Asp Leu Tyr Thr Leu Ser Ser Ser Val Thr
 195 200 205
 Val Pro Ser Ser Thr Trp Pro Ser Glu Thr Val Thr Cys Asn Val Ala
 210 215 220
 His Pro Ala Ser Ser Thr Lys Val Asp Lys Lys Ile Val Pro Arg Asp
 225 230 235 240

-continued

Cys Thr Ser Gly Gly Gly Gly Ser His His His His His His
245 250

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1644 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: Not Relevant

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Mus musculus
(B) STRAIN: BALB/c

(vii) IMMEDIATE SOURCE:
(B) CLONE: pHist22

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 88..798
(D) OTHER INFORMATION: /product= "antibody fragment, Light Chain"
/label= BotFab 22

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 818..1582
(D) OTHER INFORMATION: /product= "antibody fragment, Heavy Chain"
/label= BotFab 22

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

TATACGCCAA GCTCGAAATT AACCCCTCACT AAAGGGAACA AAAGCTGGAG CTTGAATTCT	60
TAAGTACTCG CCAAGGAGAC AGTCATA ATG AAA TAC CTA TTG CCT ACG GCG	111
Met Lys Tyr Leu Leu Pro Thr Ala	
1 5	
GCC GCT GGA TTG TTA TTA CTC GCT GCC CAA CCA GCC ATG GCC GAC ATC	159
Ala Ala Gly Leu Leu Leu Ala Ala Gln Pro Ala Met Ala Asp Ile	
10 15 20	
CAG ATG ACC CAG TCT CCA GCC TCC CTA TCT GCA TCT GTG GGA GAA ACT	207
Gln Met Thr Gln Ser Pro Ala Ser Leu Ser Ala Ser Val Gly Glu Thr	
25 30 35 40	
GTC ACT ATC ACA TGT CGA GCA AGT GGG AAT ATT CAC AAT TAT TTA GCA	255
Val Thr Ile Thr Cys Arg Ala Ser Gly Asn Ile His Asn Tyr Leu Ala	
45 50 55	
TGG TAT CAG CAG AAA CAG GGA AAA TCT CCT CAG CTC CTG GTC TAT AAT	303
Trp Tyr Gln Gln Lys Gln Gly Lys Ser Pro Gln Leu Leu Val Tyr Asn	
60 65 70	
GCA AAA ACC TTA GCA GAT GGT GTG CCA TCA AGG TTC AGT GGC AGT GGA	351
Ala Lys Thr Leu Ala Asp Gly Val Pro Ser Arg Phe Ser Gly Ser Gly	
75 80 85	
TCA GGA ACA CAA TAT TCT CTC AAG ATC AAC AGC CTG CAG CCT GAA GAT	399
Ser Gly Thr Gln Tyr Ser Leu Lys Ile Asn Ser Leu Gln Pro Glu Asp	
90 95 100	
TTT GGG AGT TAT TAC TGT CAA CAT TTT TGG AGT ACT CCG TGG ACG TTC	447
Phe Gly Ser Tyr Tyr Cys Gln His Phe Trp Ser Thr Pro Trp Thr Phe	
105 110 115 120	
GGT GGA GGC ACC AAG CTG GAA ATC AAA CGG GCT GAT GCT GCA CCA ACT	495
Gly Gly Gly Thr Lys Leu Glu Ile Lys Arg Ala Asp Ala Ala Pro Thr	
125 130 135	

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GTA	TCC	ATC	TTC	CCA	CCA	TCC	AGT	GAG	CAG	TTA	ACA	TCT	GGA	GGT	GCC	543
Val	Ser	Ile	Phe	Pro	Pro	Ser	Ser	Glu	Gln	Leu	Thr	Ser	Gly	Gly	Ala	
			140					145					150			
TCA	GTC	GTG	TGC	TTC	TTG	AAC	AAC	TTC	TAC	CCC	AAA	GAC	ATC	AAT	GTC	591
Ser	Val	Val	Cys	Phe	Leu	Asn	Asn	Phe	Tyr	Pro	Lys	Asp	Ile	Asn	Val	
		155					160					165				
AAG	TGG	AAG	ATT	GAT	GGC	AGT	GAA	CGA	CAA	AAT	GGC	GTC	CTG	AAC	AGT	639
Lys	Trp	Lys	Ile	Asp			Glu	Arg	Gln	Asn	Gly	Val	Leu	Asn	Ser	
	170					175					180					
TGG	ACT	GAT	CAG	GAC	AGC	AAA	GAC	AGC	ACC	TAC	AGC	ATG	AGC	AGC	ACC	687
Trp	Thr	Asp	Gln	Asp	Ser	Lys	Asp	Ser	Thr	Tyr	Ser	Met	Ser	Ser	Thr	
	185				190					195					200	
CTC	ACA	TTG	ACC	AAG	GAC	GAG	TAT	GAA	CGA	CAT	AAC	AGC	TAT	ACC	TGT	735
Leu	Thr	Leu	Thr	Lys	Asp	Glu	Tyr	Glu	Arg	His	Asn	Ser	Tyr	Thr	Cys	
				205					210					215		
GAG	GCC	ACT	CAC	AAG	ACA	TCA	ACT	TCA	CCC	ATT	GTC	AAG	AGC	TTC	AAC	783
Glu	Ala	Thr	His	Lys	Thr	Ser	Thr	Ser	Pro	Ile	Val	Lys	Ser	Phe	Asn	
			220					225					230			
AGG	AAT	GAG	TGT	TAA	TAGCAAGGAG	ACAGTCATA	ATG	AAA	TAC	CTA	TTG	CCT				835
Arg	Asn	Glu	Cys				Met	Lys	Tyr	Leu	Leu	Pro				
	235						1				5					
ACG	GCA	GCC	GCT	GGA	TTG	TTA	TTA	CTC	GCG	GCC	CAA	CCG	GCC	ATG	GCC	883
Thr	Ala	Ala	Ala	Gly	Leu	Leu	Leu	Ala	Ala	Gln	Pro	Ala	Met	Ala		
			10					15				20				
GAG	GTT	CAG	CTT	CAG	CAG	TCT	GGG	GCA	GAG	CTT	GTG	AAG	CCA	GGG	GCC	931
Glu	Val	Gln	Leu	Gln	Gln	Ser	Gly	Ala	Glu	Leu	Val	Lys	Pro	Gly	Ala	
		25					30					35				
TCA	GTC	AAG	TTG	TCC	TGC	ACA	GCT	TCT	GGC	TTC	AAC	ATT	AAA	GAC	ACC	979
Ser	Val	Lys	Leu	Ser	Cys	Thr	Ala	Ser	Gly	Phe	Asn	Ile	Lys	Asp	Thr	
	40					45				50						
TTT	ATG	CAC	TGG	GTG	AAG	CAG	AGG	CCT	GAA	CAG	GGC	CTG	GAG	TGG	ATT	1027
Phe	Met	His	Trp	Val	Lys	Gln	Arg	Pro	Glu	Gln	Gly	Leu	Glu	Trp	Ile	
	55				60				65					70		
GGA	AGG	ATT	GAT	CCT	GCG	AAT	GGG	AAT	ACT	GAA	TAT	GAC	CCG	AAG	TTC	1075
Gly	Arg	Ile	Asp	Pro	Ala	Asn	Gly	Asn	Thr	Glu	Tyr	Asp	Pro	Lys	Phe	
			75					80					85			
CAG	GGC	AAG	GCC	ACT	ATA	ACA	GCA	GAC	ACA	TCC	TCC	AAC	ACA	GTC	AAC	1123
Gln	Gly	Lys	Ala	Thr	Ile	Thr	Ala	Asp	Thr	Ser	Ser	Asn	Thr	Val	Asn	
		90						95					100			
CTG	CAG	CTC	AGC	AGC	CTG	ACA	TCT	GAG	GAC	ACT	GCC	GTC	TAT	TAC	TGT	1171
Leu	Gln	Leu	Ser	Ser	Leu	Thr	Ser	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	
		105				110						115				
GCT	AGT	GGA	GGG	GAA	CTG	GGG	TTT	CCT	TAC	TGG	GGC	CAA	GGG	ACT	CTG	1219
Ala	Ser	Gly	Gly	Glu	Leu	Gly	Phe	Pro	Tyr	Trp	Gly	Gln	Gly	Thr	Leu	
	120				125						130					
GTC	ACT	GTC	TCT	GCA	GCC	AAA	ACG	ACA	CCC	CCA	TCT	GTC	TAT	CCA	CTG	1267
Val	Thr	Val	Ser	Ala	Ala	Lys	Thr	Thr	Pro	Pro	Ser	Val	Tyr	Pro	Leu	
	135				140				145						150	
GCC	CCT	GGA	TCT	GCT	GCC	CAA	ACT	AAC	TCC	ATG	GTG	ACC	CTG	GGA	TGC	1315
Ala	Pro	Gly	Ser	Ala	Ala	Gln	Thr	Asn	Ser	Met	Val	Thr	Leu	Gly	Cys	
			155					160					165			
CTG	GTC	AAG	GGC	TAC	TTC	CCT	GAG	CCA	GTG	ACA	GTG	ACC	TGG	AAC	TCT	1363
Leu	Val	Lys	Gly	Tyr	Phe	Pro	Glu	Pro	Val	Thr	Val	Thr	Trp	Asn	Ser	
		170					175						180			
GGA	TCC	CTG	TCC	AGC	GGT	GTG	CAC	ACC	TTC	CCA	GCT	GTC	CTG	CAG	TCT	1411
Gly	Ser	Leu	Ser	Ser	Gly	Val	His	Thr	Phe	Pro	Ala	Val	Leu	Gln	Ser	
	185					190						195				
GAC	CTC	TAC	ACT	CTG	AGC	AGC	TCA	GTG	ACT	GTC	CCC	TCC	AGC	ACC	TGG	1459
Asp	Leu	Tyr	Thr	Leu	Ser	Ser	Ser	Val	Thr	Val	Pro	Ser	Ser	Thr	Trp	
	200					205					210					

-continued

CCC AGT GAG ACC GTC ACC TGC AAC GTT GCC CAC CCG GCC AGC AGC ACC	1507
Pro Ser Glu Thr Val Thr Cys Asn Val Ala His Pro Ala Ser Ser Thr	
215 220 225 230	
AAG GTG GAC AAG AAA ATT GTG CCC AGG GAT TGT ACT AGT GGA GGT GGA	1555
Lys Val Asp Lys Lys Ile Val Pro Arg Asp Cys Thr Ser Gly Gly Gly	
235 240 245	
GGT AGC CAC CAT CAC CAT CAC CAT TAA TCTAGAGTTA AGCGGCCGTC	1602
Gly Ser His His His His His	
250	
GAGGGGGGGC CCGGTACCCA ATTCGCCCTA TAGTGAGTCG TA	1644

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 236 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala	
1 5 10 15	
Ala Gln Pro Ala Met Ala Asp Ile Gln Met Thr Gln Ser Pro Ala Ser	
20 25 30	
Leu Ser Ala Ser Val Gly Glu Thr Val Thr Ile Thr Cys Arg Ala Ser	
35 40 45	
Gly Asn Ile His Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Gln Gly Lys	
50 55 60	
Ser Pro Gln Leu Leu Val Tyr Asn Ala Lys Thr Leu Ala Asp Gly Val	
65 70 75 80	
Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Gln Tyr Ser Leu Lys	
85 90 95	
Ile Asn Ser Leu Gln Pro Glu Asp Phe Gly Ser Tyr Tyr Cys Gln His	
100 105 110	
Phe Trp Ser Thr Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile	
115 120 125	
Lys Arg Ala Asp Ala Ala Pro Thr Val Ser Ile Phe Pro Pro Ser Ser	
130 135 140	
Glu Gln Leu Thr Ser Gly Gly Ala Ser Val Val Cys Phe Leu Asn Asn	
145 150 155 160	
Phe Tyr Pro Lys Asp Ile Asn Val Lys Trp Lys Ile Asp Gly Ser Glu	
165 170 175	
Arg Gln Asn Gly Val Leu Asn Ser Trp Thr Asp Gln Asp Ser Lys Asp	
180 185 190	
Ser Thr Tyr Ser Met Ser Ser Thr Leu Thr Leu Thr Lys Asp Glu Tyr	
195 200 205	
Glu Arg His Asn Ser Tyr Thr Cys Glu Ala Thr His Lys Thr Ser Thr	
210 215 220	
Ser Pro Ile Val Lys Ser Phe Asn Arg Asn Glu Cys	
225 230 235	

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 254 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

-continued

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala
 1 5 10 15
 Ala Gln Pro Ala Met Ala Glu Val Gln Leu Gln Gln Ser Gly Ala Glu
 20 25 30
 Leu Val Lys Pro Gly Ala Ser Val Lys Leu Ser Cys Thr Ala Ser Gly
 35 40 45
 Phe Asn Ile Lys Asp Thr Phe Met His Trp Val Lys Gln Arg Pro Glu
 50 55 60
 Gln Gly Leu Glu Trp Ile Gly Arg Ile Asp Pro Ala Asn Gly Asn Thr
 65 70 75 80
 Glu Tyr Asp Pro Lys Phe Gln Gly Lys Ala Thr Ile Thr Ala Asp Thr
 85 90 95
 Ser Ser Asn Thr Val Asn Leu Gln Leu Ser Ser Leu Thr Ser Glu Asp
 100 105 110
 Thr Ala Val Tyr Tyr Cys Ala Ser Gly Gly Glu Leu Gly Phe Pro Tyr
 115 120 125
 Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ala Ala Lys Thr Thr Pro
 130 135 140
 Pro Ser Val Tyr Pro Leu Ala Pro Gly Ser Ala Ala Gln Thr Asn Ser
 145 150 155 160
 Met Val Thr Leu Gly Cys Leu Val Lys Gly Tyr Phe Pro Glu Pro Val
 165 170 175
 Thr Val Thr Trp Asn Ser Gly Ser Leu Ser Ser Gly Val His Thr Phe
 180 185 190
 Pro Ala Val Leu Gln Ser Asp Leu Tyr Thr Leu Ser Ser Ser Val Thr
 195 200 205
 Val Pro Ser Ser Thr Trp Pro Ser Glu Thr Val Thr Cys Asn Val Ala
 210 215 220
 His Pro Ala Ser Ser Thr Lys Val Asp Lys Lys Ile Val Pro Arg Asp
 225 230 235 240
 Cys Thr Ser Gly Gly Gly Gly Ser His His His His His His
 245 250

We claim:

1. A purified and isolated recombinant antibody fragment (rFab) comprising a light chain polypeptide and a heavy chain polypeptide, each of said polypeptides having an N-terminus and a carboxyl terminus, wherein said light chain polypeptide consists of the amino acid sequence of SEQ ID NO:3 and said heavy chain polypeptide consists of the amino acid sequence of SEQ ID NO:4, and wherein said rFab is capable of binding specifically to the non-neurotoxin component of a botulinum neurotoxin complex selected from the group consisting of type A and type B.

2. The rFab of claim 1, further comprising a six amino acid histidine tail fused to said carboxyl terminus of said heavy chain polypeptide.

3. A purified and isolated DNA fragment encoding an amino acid sequence for a recombinant antibody fragment, said antibody fragment comprising a light chain polypeptide and a heavy chain polypeptide, wherein the light chain polypeptide consists of the amino acid sequence of SEQ ID NO:3 and the heavy chain polypeptide consists of the amino acid sequence of SEQ ID NO:4, and wherein said purified and isolated DNA fragment comprises a first DNA sequence coding for the light chain polypeptide and a second DNA sequence coding for the heavy chain polypeptide.

4. The DNA fragment of claim 3, wherein said first DNA sequence comprises nucleotides 117-827 of SEQ ID NO:2 and said second DNA sequence comprises nucleotides 847-1611 of SEQ ID NO:2.

5. The DNA fragment of claim 4, further comprising a third DNA sequence located between said first and second DNA sequences, said third DNA sequence consisting of nucleotides 828-846 of SEQ ID NO:2.

6. The DNA fragment of claim 3, wherein said first DNA sequence comprises the complementary sequence of nucleotides 117-827 of SEQ ID NO:2 and said second DNA sequence comprises the complementary sequence of nucleotides 847-1611 of SEQ ID NO:2.

7. The DNA fragment of claim 6, further comprising a third DNA sequence located between said first and second DNA sequences, said third DNA sequence consisting of nucleotides 828-846 of SEQ ID NO:2.

8. A purified and isolated recombinant antibody fragment (rFab) comprising a light chain polypeptide and a heavy chain polypeptide, each of said polypeptides having an N-terminus and a carboxyl terminus, wherein said light chain polypeptide consists of the amino acid sequence of SEQ ID NO:6 and said heavy chain polypeptide consists of the amino acid sequence of SEQ ID NO:7, and wherein said

rFab is capable of binding specifically to the non-neurotoxin component of a botulinum neurotoxin complex selected from the group consisting of type A and type B.

9. The rFab of claim 8, further comprising a six amino acid histidine tail fused to said carboxyl terminus of said heavy chain polypeptide.

10. A purified and isolated DNA fragment encoding an amino acid sequence for a recombinant antibody fragment, said antibody fragment comprising a light chain polypeptide and a heavy chain polypeptide, wherein said light chain polypeptide consists of the amino acid sequence of SEQ ID NO:6 and the heavy chain polypeptide consists of the amino acid sequence of SEQ ID NO:7, and wherein said purified and isolated DNA fragment comprises a first DNA sequence coding for the light chain polypeptide and a second DNA sequence coding for the heavy chain polypeptide.

11. The DNA fragment of claim 10, wherein said first DNA sequence comprises nucleotides 87–788 of SEQ ID NO:5 and said second DNA sequence comprises nucleotides 815–1579 of SEQ ID NO:5.

12. The DNA fragment of claim 11, further comprising a third DNA sequence located between said first and second DNA sequences, said third DNA sequence consisting of nucleotides 789–814 of SEQ ID NO:5.

13. The DNA fragment of claim 10, wherein said first DNA sequence comprises the complementary sequence of nucleotides 87–788 of SEQ ID NO:5 and said second DNA sequence comprises the complementary sequence of nucleotides 815–1579 of SEQ ID NO:5.

14. The DNA fragment of claim 13, further comprising a third DNA sequence located between said first and second DNA sequences, said third DNA sequence consisting of nucleotides 789–814 of SEQ ID NO:5.

15. A purified and isolated recombinant antibody fragment (rFab) comprising a light chain polypeptide and a heavy chain polypeptide, each of said polypeptides having an N-terminus and a carboxyl terminus, wherein said light chain polypeptide consists of the amino acid sequence of SEQ ID NO:9 and said heavy chain polypeptide consists of the amino acid sequence of SEQ ID NO:10, and wherein said rFab is capable of binding specifically to the non-neurotoxin component of a botulinum neurotoxin complex selected from the group consisting of type A and type B.

16. The rFab of claim 15, further comprising a six amino acid histidine tail fused to said carboxyl terminus of said heavy chain polypeptide.

17. A purified and isolated DNA fragment encoding an amino acid sequence for a recombinant antibody fragment, said antibody fragment comprising a light chain polypeptide and a heavy chain polypeptide, wherein said light chain polypeptide consists of the amino acid sequence of SEQ ID NO:9 and the heavy chain polypeptide consists of the amino acid sequence of SEQ ID NO:10, and wherein said purified and isolated DNA fragment comprises a first DNA sequence coding for the light chain polypeptide and a second DNA sequence coding for the heavy chain polypeptide.

18. The DNA fragment of claim 17, wherein said first DNA sequence comprises nucleotides 88–798 of SEQ ID NO:8 and said second DNA sequence comprises nucleotides 818–1582 of SEQ ID NO:8.

19. The DNA fragment of claim 18, further comprising a third DNA sequence located between said first and second DNA sequences, said third DNA sequence consisting of nucleotides 799–817 of SEQ ID NO:8.

20. The DNA fragment of claim 17, wherein said first DNA sequence comprises the complementary sequence of nucleotides 88–798 of SEQ ID NO:8 and said second DNA sequence comprises the complementary sequence of nucleotides 818–1582 of SEQ ID NO:8.

21. The DNA fragment of claim 20, further comprising a third DNA sequence located between said first and second

DNA sequences, said third DNA sequence consisting of nucleotides 799–817 of SEQ ID NO:8.

22. A purified and isolated recombinant antibody fragment (rFab) comprising a light chain polypeptide and a heavy chain polypeptide, each of said polypeptides having an N-terminus and a carboxyl terminus, wherein said light chain polypeptide consists of the amino acid sequence of SEQ ID NO:12 and said heavy chain polypeptide consists of the amino acid sequence of SEQ ID NO:13, and wherein said rFab is capable of binding specifically to the non-neurotoxin component of a botulinum neurotoxin complex selected from the group consisting of type A and type B.

23. The rFab of claim 22, further comprising a six amino acid histidine tail fused to said carboxyl terminus of said heavy chain polypeptide.

24. A purified and isolated DNA fragment encoding an amino acid sequence for a recombinant antibody fragment, said antibody fragment comprising a light chain polypeptide and a heavy chain polypeptide, wherein said light chain polypeptide consists of the amino acid sequence of SEQ ID NO:12 and the heavy chain polypeptide consists of the amino acid sequence of SEQ ID NO:13, and wherein said purified and isolated DNA fragment comprises a first DNA sequence coding for the light chain polypeptide and a second DNA sequence coding for the heavy chain polypeptide.

25. The DNA fragment of claim 24, wherein said first DNA sequence comprises nucleotides 88–798 of SEQ ID NO:11 and said second DNA sequence comprises nucleotides 818–1582 of SEQ ID NO:11.

26. The DNA fragment of claim 25, further comprising a third DNA sequence located between said first and second DNA sequences, said third DNA sequence consisting of nucleotides 799–817 of SEQ ID NO:11.

27. The DNA fragment of claim 24, wherein said first DNA sequence comprises the complementary sequence of nucleotides 88–798 of SEQ ID NO:11 and said second DNA sequence comprises the complementary sequence of nucleotides 818–1582 of SEQ ID NO:11.

28. The DNA fragment of claim 27, further comprising a third DNA sequence located between said first and second DNA sequences, said third DNA sequence consisting of nucleotides 799–817 of SEQ ID NO:11.

29. A recombinant expression vector for expressing, in a microorganism, an rFab against the non-neurotoxin component of botulinum complex type A or type B, said rFab having a light chain polypeptide and a heavy chain polypeptide, said expression vector comprising:

- (a) a first DNA sequence which codes for said light chain polypeptide, said polypeptide having an N-terminus and a carboxyl terminus;
- (b) a second DNA sequence which codes for said heavy chain polypeptide, said polypeptide having an N-terminus and a carboxyl terminus; and
- (c) transcriptional and translational control elements recognized by said microorganism which are operatively linked to said first and second DNA sequences; and wherein said expression vector comprises the nucleotide sequence of SEQ ID NO:1.

30. The expression vector of claim 29, further comprising a third DNA sequence coding for a histidine tail fused to the carboxyl terminus of said second DNA sequence.

31. A recombinant microorganism which expresses an rFab against the non-neurotoxin component of botulinum neurotoxin complex type A or type B, wherein the microorganism contains a recombinant expression vector which comprises the nucleotide sequence of SEQ ID NO:1.

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